Supplementary Table S1. Strains of *Acinetobacter beijerinckii* sp. nov. and *Acinetobacter gyllenbergii* sp. nov.

CCM, Czech Collection of Microorganisms, Brno, Czech Republic; CCUG, Culture Collection, University of Göteborg, Sweden; LUH and RUH, Collection L. Dijkshoorn, Leiden University Medical Center, Leiden, the Netherlands; NIPH, Collection A. Nemec, National Institute of Public Health, Prague, Czech Republic; in, inpatient; out, outpatient.

Strain designation	Specimen	Location and year of isolation	Donator	Reference	16S rRNA gene EMBL accession no.	rpoB gene GenBank accession no.
Acinetobacter beijerinckii (*n*=15)						
NIPH 838^{T} (= LUH 4759^{T} = 58a^{T}† = CCM 7266^{T} = CCUG 51249^{T})	Wound (human, in)	Malmö, Sweden, 1980	I. Tjernberg	Tjernberg & Ursing (1989)	AJ626712	EU477124
LUH 6214 (= NIPH 1453 = DVL00/879‡ = CCUG 56139)	Airsacculitis (horse)	Gent, Belgium, 2000	L.A. Devriese		AJ303013	EU477130
LUH 4738 (= NIPH 832 = 83†)	Peritoneal dialysis fluid (human)	Unknown	P.J.M. Bouvet	Bouvet & Grimont (1986)		EU477123
LUH 4561 (= NIPH 770 = A136b‡)	Soil from footpath	Pilion Peninsula, Greece, 1993-1994	H. Seifert			EU477120
LUH 3340 (= NIPH 1065 = SH 139†)	Toe-web (human)	Cologne, Germany, 1994	H. Seifert	Seifert *et al.* (1997)		EU477129
LUH 4771 (= NIPH 850 = 190†)	Plastic foam	Malmö, Sweden, 1981	I. Tjernberg	Tjernberg & Ursing (1989)		EU477125
RUH 2371 (= NIPH 2011)	Sputum (human, in)	Rotterdam, the Netherlands, 1987				EU477137
RUH 2560 (= NIPH 2013)	Feces (human, in)	Rotterdam, the Netherlands, 1987				EU477138
RUH 2762 (= NIPH 2014)	Throat (human, in)	Rotterdam, the Netherlands, 1988				EU477139
RUH 2879 (= NIPH 2015)	Gall (human, in)	Rotterdam, the Netherlands, 1988				EU477140
LUH 3146 (= NIPH 2016 = 1295-124-2‡)	Skin of hospital staff	Enschede, the Netherlands, 1995	A. Bernards			EU477141
LUH 7834 (= NIPH 2017 = v0202228‡)	Airways infection (horse)	Lelystadt, the Netherlands, 2002	J. Wagenaar			EU477142
LUH 8896 (= NIPH 2018 = 03B041980‡)	Sputum (human)	Leeuwarden, the Netherlands, 2003	J. van Zeijl			EU477143
LUH 5692 (= NIPH 2025 = VMDC 9702631‡)	Surface water	Utrecht, the Netherlands, 1999	J. Wagenaar			EU477146
LUH 9424 (= NIPH 2372 = CCUG 56141)	Perineum (human, in)	Leiden, the Netherlands, 2000				EU477157
Acinetobacter gyllenbergii (*n*=9)						
NIPH 2150^{T} (= RUH 422^{T} = 1271^{T}† = CCM 7267^{T} = CCUG 51248^{T})	Urine (human)	Leiden, the Netherlands, 1978	P.J.M. Bouvet	Bouvet & Jeanjean (1989)	AJ293694	EU477148
LUH 1740 (= NIPH 975 = 930†)	Tracheal exudate (human)	France	P.J.M. Bouvet	Bouvet & Jeanjean (1989)	AJ293693	EU477127
LUH 1737 (= NIPH 802 = 80†)	Blood (human)	France	P.J.M. Bouvet	Bouvet & Jeanjean (1989)	AJ293692	EU477121
LUH 1741 (= NIPH 822 = 944†)	Wound (human)	France	P.J.M. Bouvet	Bouvet & Jeanjean (1989)		EU477158
NIPH 230 (= LUH 4712 = CCUG 56138)	Vagina (human, out)	Praha, Czech Republic, 1994		Nemec *et al.* (2000)		EU477106
LUH 5809 (= NIPH 1773 = 3268‡)	Tracheal aspirate (human, in)	Hong Kong, China, 1998	E.T. Houang			EU477131
LUH 6541 (= NIPH 2021 = CCUG 56140)	Sinus (human)	Leiden, the Netherlands, 2000				EU477144
RUH 3064 (= NIPH 2022)	Sputum (human, in)	Rotterdam, the Netherlands, 1989				EU477145
NIPH 2353 (= LUH 9706 = CCM 1976)	Throat (human)	Czech Republic	CCM	Lehmann (1973)		EU477156

† Strain designation used in a reference publication.

‡ Strain designation used by a donator.

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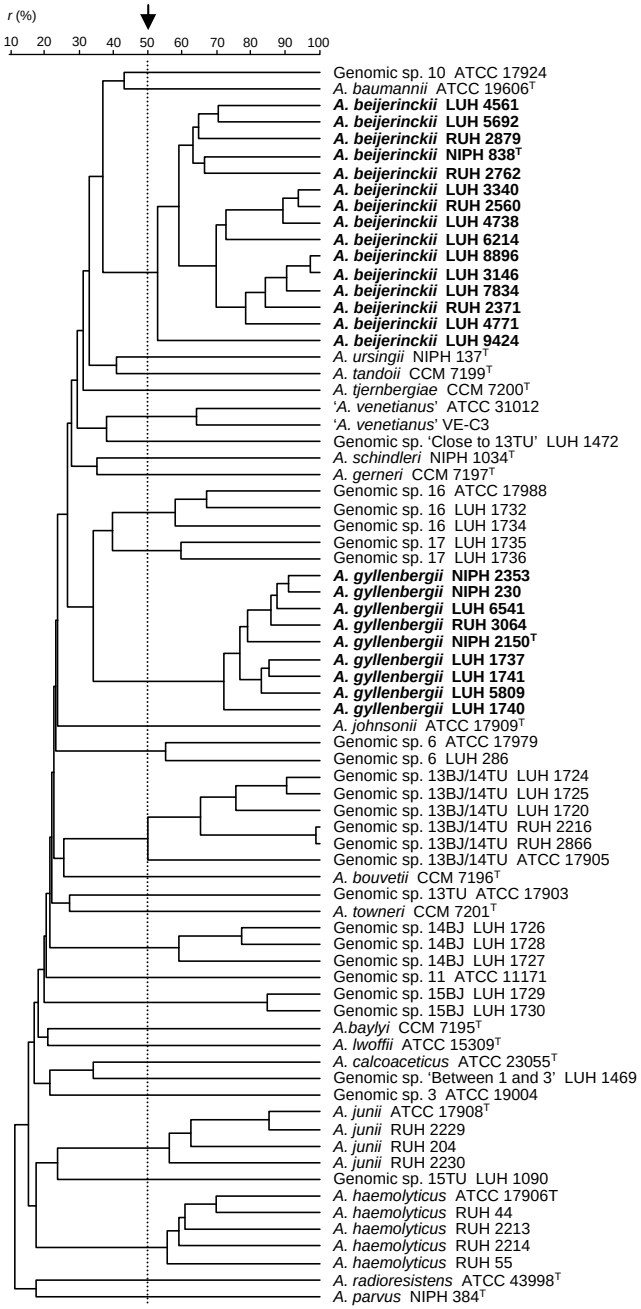

Supplementary Fig. S1. Dendrogram of cluster analysis of AFLP fingerprints of 15 strains of *Acinetobacter beijerinckii* sp. nov., 9 strains of *Acinetobacter gylenbergii* sp. nov., and 50 strains representing all known genomic species of the genus *Acinetobacter*. Species associated with haemolytic activity are represented by multiple strains, which were previously studied by DNA–DNA hybridization; detailed information on these strains can be found in the studies of Janssen *et al*. [*Int J Syst Bacteriol* (1997) **47**, 1179–1187) and Vaneechoutte *et al*. [*Res Microbiol* (1999) **150**, 69–73]. AFLP was performed as described [Nemec *et al*. *Int J Syst Evol Microbiol* (2001) **51**, 1891–1899], with the following steps: simultaneous digestion of DNA with two restriction endonucleases (*Eco*RI and *Mse*I) and adapter ligation, PCR with a Cy5-labelled *Eco*RI+A primer and a *Mse*I+C primer (A, C = selective nucleotides), separation of amplified fragments with the ALF Express system (Amersham Biosciences) and cluster analysis of patterns with BioNumerics software release 4.6 (Applied Maths) using Pearson's product (*r*) for similarity calculation and UPGMA for clustering. The arrow indicates the species delineation level.